

Invasion of a holarctic planktonic cladoceran daphnia galeata sars (Crustacea: Cladocera) in the lower lakes of South Australia

Kazan Federal University, 420008, Kremlevskaya 18, Kazan, Russia

Abstract

Copyright © 2018 Magnolia Press. We found a Holarctic microcrustacean *Daphnia galeata* Sars, 1863 (Cladocera: Daphniidae) in the Lower Lakes of South Australia. This taxon was never detected in continental Australia before. Its identity was confirmed by the sequences of mitochondrial COI, 12S and 16S and nuclear 18S and 28S genes. A maximum likelihood tree from a dataset from combining 12S + 16S mitochondrial sequence and a split network of the COI haplotypes are provided, but resolution of both genes is not sufficient to reveal the exact region of the Holarctic from where *D. galeata* was introduced to Australia; the vector of its invasion also is unknown. We hypothesize that appearance of *D. galeata* in the Lower Lakes of the Murray River is related to a recent anthropogenic eutrophication of water bodies in this region, keeping in mind that examples of successful invasion of some European lakes by *D. galeata* after their eutrophication are well-known. We also hypothesize that establishment of this non-indigenous taxon populations in Australia might have a strong negative impact on native lake biota.

<http://dx.doi.org/10.11646/zootaxa.4402.1.6>

Keywords

Australia, Biological invasion, Cladocera, Daphnia, Eutrophication

References

- [1] Adamowicz, S.J., Petrusek, A., Colbourne, J.K., Hebert, P.D.N. & Witt, J.D.S. (2009) The scale of divergence: a phylogenetic appraisal of intercontinental allopatric speciation in a passively dispersed freshwater zooplankton genus. *Molecular Phylogenetics and Evolution*, 50, 423–436. <https://doi.org/10.1016/j.ympev.2008.11.026>
- [2] Allen, G.R., Midgley, S.H. & Allen, M. (2002) *Field Guide to the Freshwater Fishes of Australia*. Western Australian Museum, Perth, 410 pp.
- [3] Balvert, S.F., Duggan, I.C. & Hogg, I.D. (2009) Zooplankton seasonal dynamics in a recently filled mine pit lake: the effect of nonindigenous *Daphnia* establishment. *Aquatic Ecology*, 43, 403–413. <https://doi.org/10.1007/s10452-008-9165-z>
- [4] Bekker, E.I., Karabanov, D.P., Galimov, Y.R. & Kotov, A.A. (2016) DNA barcoding reveals high cryptic diversity in the North Eurasian *Moina* species (Crustacea: Cladocera). *PLoS ONE*, 11 (8), e0161737. <https://doi.org/10.1371/journal.pone.0161737>
- [5] Belyaeva, M. & Taylor, D.J. (2009) Cryptic species within the *Chydorus sphaericus* species complex (Crustacea: Cladocera) revealed by molecular markers and sexual stage morphology. *Molecular Phylogenetics and Evolution*, 50, 534–546. <https://doi.org/10.1016/j.ympev.2008.11.007>

- [6] Benzie, J.A. (2005) The Genus *Daphnia* (including *Daphniopsis*): (Anomopoda, Daphniidae). Vol. 21. Backhuys Publishers, Leiden and Kenobi Productions, Ghent, vii + 376 pp.
- [7] Benzie, J.A.H. & Hodges, A.M.A. (1996) *Daphnia obtusa* Kurz, 1874 emend Scourfield, 1942 from Australia. *Hydrobiologia*, 333, 195–199. <https://doi.org/10.1007/BF00013433>
- [8] Bouckaert, R., Heled, J., Kuhnert, D., Vaughan, T., Wu, C.H., Xie, D., Suchard, M.A., Rambaut, A. & Drummond, A.J. (2014) BEAST 2: A Software Platform for Bayesian Evolutionary Analysis. *PLoS Computational Biology*, 10 (4), e1003537. <https://doi.org/10.1371/journal.pcbi.1003537>
- [9] Brede, N., Sandrock, C., Straile, D., Spaak, P., Jankowski, T., Streit, B. & Schwenk, K. (2009) The impact of human-made ecological changes on the genetic architecture of *Daphnia* species. *Proceedings of the National Academy of Sciences*, 106, 4758–4763. <https://doi.org/10.1073/pnas.0807187106>
- [10] Briski, E., Cristescu, M.E., Bailey, S.A. & Macisaac, H.J. (2011) Use of DNA barcoding to detect invertebrate invasive species from diapausing eggs. *Biological Invasions*, 13, 1325–1340. <https://doi.org/10.1007/s10530-010-9892-7>
- [11] Briski, E., Bailey, S.A., Casas-Monroy, O., DiBacco, C., Kaczmarska, I., Lawrence, J.E., Leichsenring, J., Levings, C., MacGillivray, M.L., McKindsey, C.W., Nasmith, L.E., Parenteau, M., Piercey, G.E., Rivkin, R.B., Rochon, A., Roy, S., Simard, N., Sun, B., Way, C., Weise, A.M. & MacIsaac, H.J. (2013) Taxon and vector specific variation in species richness and abundance during the transport stage of biological invasions. *Limnology and Oceanography*, 58 (4), 1361–1372. <https://doi.org/10.4319/lo.2013.58.4.1361>
- [12] Brooks, J.L. (1957) The systematics of North American *Daphnia*. *Memoirs of the Connecticut Academy of Arts and Sciences*, 13, 1–180.
- [13] Burns, C.W. (2013) Predictors of invasion success by *Daphnia* species: influence of food, temperature and species identity. *Biological Invasions*, 15, 859–869. <https://doi.org/10.1007/s10530-012-0335-5>
- [14] Chapman, M.A., Lewis, M.H. & Stout, V.M. (1976) Introduction to the freshwater crustacea of New Zealand. Collins, Auckland, 261 pp.
- [15] Colbourne, J.K., Wilson, C.C. & Hebert, P.D.N. (2006) The systematics of Australian *Daphnia* and *Daphniopsis* (Crustacea: Cladocera): a shared phylogenetic history transformed by habitat-specific rates of evolution. *Biological Journal of the Linnean Society*, 89, 469–488. <https://doi.org/10.1111/j.1095-8312.2006.00687.x>
- [16] Crease, T.J., Omilian, A.R., Costanzo, K.S. & Taylor, D.J. (2012) Transcontinental phylogeography of the *Daphnia pulex* species complex. *PLoS ONE*, 7 (10), e46620. <https://doi.org/10.1371/journal.pone.0046620>
- [17] Colbourne, J.K. & Hebert, P.D.N. (1996) The systematics of North American *Daphnia* (Crustacea: Anomopoda): a molecular phylogenetic approach. *Philosophical Transactions Royal Society, Series B*, 351, 349–360. <https://doi.org/10.1098/rstb.1996.0028>
- [18] Darriba, D., Taboada, G., Doallo, R. & Posada, D. (2012) jModelTest 2: more models, new heuristics and parallel computing. *Nature Methods*, 9, 772. <https://doi.org/10.1038/nmeth.2109>
- [19] Duggan, I.C., Green, J.D. & Burger, D.F. (2006) First New Zealand records of three non-indigenous zooplankton species: *Skistodiaptomus pallidus*, *Sinodiaptomus valkanovi*, and *Daphnia dentifera*. *New Zealand Journal of Marine and Freshwater Research*, 40, 561–569. <https://doi.org/10.1080/00288330.2006.9517445>
- [20] Duggan, I.C. & Pullan, S.G. (2017) Do freshwater aquaculture facilities provide an invasion risk for zooplankton hitchhikers. *Biological Invasions*, 19, 307–314. <https://doi.org/10.1007/s10530-016-1280-5>
- [21] Duggan, I.C., Robinson, K.V., Burns, C.W., Banks, J. & Hogg, I. (2012) Identifying invertebrate invasions using morphological and molecular analyses: North American *Daphnia* 'pulex' in New Zealand fresh waters. *Aquatic Invasions*, 7, 585–590. <https://doi.org/10.3391/ai.2012.7.4.015>
- [22] Elton, C.S. (1958) *The Ecology of Invasions by Animals and Plants*. Foreword by Daniel Simberloff. University of Chicago Press, Chicago, 196 pp. <https://doi.org/10.1007/978-1-4899-7214-9>
- [23] Forró, L., Korovchinsky, N.M., Kotov, A.A. & Petrusek, A. (2008) Global diversity of cladocerans (Cladocera; Crustacea) in freshwater. *Hydrobiologia*, 595, 177–184. <https://doi.org/10.1007/s10750-007-9013-5>
- [24] Geddes, M.C. (1984) Seasonal studies on the zooplankton community of Lake Alexandrina, River Murray, South Australia, and the role of turbidity in determining zooplankton community structure. *Marine and Freshwater Research*, 35 (4), 417–426. <https://doi.org/10.1071/MF9840417>
- [25] Geddes, M.C., Shiel, R.J. & Francis, J. (2016) Zooplankton in the Murray estuary and Coorong during flow and no-flow periods. *Transactions of the Royal Society of South Australia*, 140, 74–89. <https://doi.org/10.1080/03721426.2016.1151497>
- [26] Geller, J., Meyer, C., Parker, M. & Hawk, H. (2013) Redesign of PCR primers for mitochondrial cytochrome c oxidase subunit I for marine invertebrates and application in all-taxa biotic surveys. *Molecular Ecology Resources*, 13, 851–861. <https://doi.org/10.1111/1755-0998.12138>
- [27] Giessler, S. & Englbrecht, C.C. (2009) Dynamic reticulate evolution in a *Daphnia* multispecies complex. *Journal of Experimental Zoology*, 311A, 531–549. <https://doi.org/10.1002/jez.550>
- [28] Gruber, A.R., Bernhart, S.H. & Lorenz, R. (2015) The ViennaRNA web services. *Methods in Molecular Biology*, 1269, 307–326. https://doi.org/10.1007/978-1-4939-2291-8_19

- [29] Hairston, N.G.Jr., Holtmeier, C.L., Lampert, W., Weider, L.J., Post, D.M., Fischer, J.M., Cáceres, C.E., Fox, J.A. & Gaedke, U. (2001) Natural selection for grazer resistance to toxic cyanobacteria: Evolution of phenotypic plasticity. *Evolution*, 55, 2203–2214. <https://doi.org/10.1111/j.0014-3820.2001.tb00736.x>
- [30] Hebert, P.D.N., Cywinska, A., Ball, S.L. & De Waard, J.R. (2003) Biological identifications through DNA barcodes. *Proceedings of the Royal Society B: Biological Sciences*, 270, 313–321. <https://doi.org/10.1098/rspb.2002.2218>
- [31] Horwitz, P. (1990) The translocation of freshwater crayfish in Australia: potential impact, the need for control and global relevance. *Biological Conservation*, 54, 291–305. [https://doi.org/10.1016/0006-3207\(90\)90142-C](https://doi.org/10.1016/0006-3207(90)90142-C)
- [32] Huson, D.H., Rupp, R. & Scornavacca, C. (2010) *Phylogenetic Networks: Concepts, Algorithms and Applications*. Cambridge University Press, Cambridge, 376 pp. <https://doi.org/10.1017/CBO9780511974076>
- [33] Incagnone, G., Marrone, F., Barone, R., Robba, L. & Naselli-Flores, K. (2014) How do freshwater organisms cross the “dry ocean”. A review on passive dispersal and colonization processes with a special focus on temporary ponds. *Hydrobiologia*, 750, 103–123. <https://doi.org/10.1007/s10750-014-2110-3>
- [34] Ishida, S. & Taylor, D.J. (2007) Quaternary diversification in a sexual Holarctic zooplankter, *Daphnia galeata*. *Molecular Ecology*, 16, 569–582. <https://doi.org/10.1111/j.1365-294X.2006.03160.x>
- [35] Jankowski, T. & Straile, D. (2003) A comparison of egg-bank and long-term plankton dynamics of two *Daphnia* species, *D. hyalina* and *D. galeata*: Potentials and limits of reconstruction. *Limnology and Oceanography*, 48, 1948–1955. <https://doi.org/10.4319/lo.2003.48.5.1948>
- [36] Johnson, M., Zaretskaya, I., Raytselis, Y., Merezukh, Y., McGinnis, S. & Madden, T.L. (2008) NCBI BLAST: a better web interface. *Nucleic Acids Research*, 36, W5–W9. <https://doi.org/10.1093/nar/gkn201>
- [37] Kimura, M. (1980) A simple method for estimating evolutionary rate of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution*, 16, 111–120. <https://doi.org/10.1007/BF01731581>
- [38] Koehn, J.D., Brumley, A.R. & Gehrke, P.C. (2000) *Managing the impacts of carp*. Bureau of Rural Sciences, Canberra, 249 pp.
- [39] Koenemann, S., Jenner, R.A., Hoenemann, M., Stemme, T. & von Reumont, B.M. (2010) Arthropod phylogeny revisited, with a focus on crustacean relationships. *Arthropod Structure & Development*, 39, 88–110. <https://doi.org/10.1016/j.asd.2009.10.003>
- [40] Kotov, A.A. (2015) A critical review of the current taxonomy of the genus *Daphnia* O. F. Müller, 1785. *Zootaxa*, 3911 (2), 184–200. <https://doi.org/10.11646/zootaxa.3911.2.2>
- [41] Kotov, A.A., Karabanov, D.P., Bekker, E.I., Neretina, T.V. & Taylor, D.J. (2016) Phylogeography of the *Chydorus sphaericus* group (Cladocera: Chydoridae) in the Northern Palearctic. *PLoS ONE*, 11 (12), e0168711. <https://doi.org/10.1371/journal.pone.0168711>
- [42] Kotov, A.A. & Taylor, D.J. (2010) A new African lineage of the *Daphnia obtusa* group (Cladocera: Daphniidae) disrupts continental vicariance patterns. *Journal of Plankton Research*, 32, 937–949. <https://doi.org/10.1093/plankt/fbq018>
- [43] Kotov, A.A. & Taylor, D.J. (2014) *Daphnia lumholtzi* Sars, 1885 (Cladocera: Daphniidae) invades Argentina. *Journal of Limnology*, 73, 167–172. <https://doi.org/10.4081/jlimnol.2014.920>
- [44] Kumar, S., Stecher, G. & Tamura, K. (2016) MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution*, 33, 1870–1874. <https://doi.org/10.1093/molbev/msw054>
- [45] Li, S., Bush, R., Mao, R., Xiong, L. & Ye, C. (2017) Extreme drought causes distinct water acidification and eutrophication in the Lower Lakes (Lakes Alexandrina and Albert), Australia. *Journal of Hydrology*, 544, 133–146. <https://doi.org/10.1016/j.jhydrol.2016.11.015>
- [46] Loo, S.E., Mac Nally, R. & Lake, P.S. (2007) Forecasting the range of invasion of the New Zealand mudsnail: A comparison of models built with native and invaded range data. *Ecological Applications*, 17, 181–189. [https://doi.org/10.1890/1051-0761\(2007\)017\[0181:FNZMIR\]2.0.CO;2](https://doi.org/10.1890/1051-0761(2007)017[0181:FNZMIR]2.0.CO;2)
- [47] Ma, X., Petrusek, A., Wolinska, J., Giessler, S., Zhong, Y., Yang, Z., Hu, W. & Yin, M. (2015) Diversity of the *Daphnia longispina* species complex in Chinese lakes: a DNA taxonomy approach. *Journal of Plankton Research*, 371, 55–65. <https://doi.org/10.1093/plankt/fbu091>
- [48] Marohasy, J. & Abbot, J. (2012) Deconstructing the native fish strategy for Australia’s Murray Darling catchment. *River Basin Management VII*, 172, 339.
- [49] Mergeay, J., Verschuren, D. & De Meester, L. (2005) Cryptic invasion and dispersal of an American *Daphnia* in East Africa. *Limnology & Oceanography*, 50, 1278–1283. <https://doi.org/10.4319/lo.2005.50.4.1278>
- [50] Morgan, D.L., Gill, H.S., Maddern, M.G. & Beatty, S.J. (2004) Distribution and impacts of introduced freshwater fishes in Western Australia. *New Zealand Journal of Marine and Freshwater Research*, 38, 511–523. <https://doi.org/10.1080/00288330.2004.9517257>
- [51] Mosley, L.M., Zammit, B., Leyden, E., Heneker, T.M., Hipsey, M.R., Skinner, D. & Aldridge, K.T. (2012) The impact of extreme low flows on the water quality of the Lower Murray River and Lakes (South Australia). *Water Resource Management*, 26, 3923–3946. <https://doi.org/10.1007/s11269-012-0113-2>

- [52] Okonechnikov, K., Golosova, O. & Fursov, M. (2012) Unipro UGENE: a unified bioinformatics toolkit. *Bioinformatics*, 28, 1166–1167. <https://doi.org/10.1093/bioinformatics/bts091>
- [53] Ozdemir, E., Altindag, A. & Kandemir, I. (2016) Molecular diversity of some species belonging to the genus *Daphnia* O. F. Muller, 1785 (Crustacea: Cladocera) in Turkey. *Mitochondrial DNA Part A*, 28, 424–433. <https://doi.org/10.3109/19401736.2015.1136303>
- [54] Petrusek, A., Hobaek, A., Nilssen, J.P., Skage, M., Černý, M., Brede, N. & Schwenk, K. (2008) A taxonomic reappraisal of the European *Daphnia longispina* complex (Crustacea, Cladocera, Anomopoda). *Zoologica Scripta*, 37, 507–519. <https://doi.org/10.1111/j.1463-6409.2008.00336.x>
- [55] Petrusek, A., Tollrian, R., Schwenk, K., Haas, A. & Laforsch, C. (2009) A “crown of thorns” is an inducible defense that protects *Daphnia* against an ancient predator. *Proceedings of the National Academy of Sciences USA*, 106, 2248–2252. <https://doi.org/10.1073/pnas.0808075106>
- [56] Popova, E.Y. & Kotov, A.A. (2013) Latitudinal patterns in the diversity of two subgenera of the genus *Daphnia* O.F. Müller (Crustacea: Cladocera: Daphniidae). *Zootaxa*, 3736 (2), 159–174. <https://doi.org/10.11646/zootaxa.3736.2.4>
- [57] Posada, D. & Buckley, T. (2004) Model selection and model averaging in phylogenetics: advantages of Akaike Information Criterion and Bayesian approaches over likelihood ratio tests. *Systematic Biology*, 53, 793–808. <https://doi.org/10.1080/10635150490522304>
- [58] Rellstab, C., Keller, B., Girardclos, S., Anselmetti, F.S. & Spaak, P. (2011) Anthropogenic eutrophication shapes the past and present taxonomic composition of hybridizing *Daphnia* in unproductive lakes. *Limnology and Oceanography*, 56, 292–302. <https://doi.org/10.4319/lo.2011.56.1.0292>
- [59] Reumont, B.M., Meusemann, K., Szucsich, N.U., Dell'Ampio, E., Gowri-Shankar, V., Bartel, D., Simon, S., Letsch, H.O., Stocsits, R.R., Luan, Y.X., Wägele, J.W., Pass, G., Hadrys, H. & Misof, B. (2009) Can comprehensive background knowledge be incorporated into substitution models to improve phylogenetic analyses. A case study on major arthropod relationships. *BMC Evolutionary Biology*, 9, 119. <https://doi.org/10.1186/1471-214-9-119>
- [60] Reynolds, C., Miranda, N.A. & Cumming, G.S. (2015) The role of waterbirds in the dispersal of aquatic alien and invasive species. *Diversity and Distributions*, 21, 744–754. <https://doi.org/10.1111/ddi.12334>
- [61] Rodriguez, F., Oliver, J., Marin, A. & Medina, J. (1990) The general stochastic model of nucleotide substitution. *Journal of Theoretical Biology*, 142, 485–501. [https://doi.org/10.1016/S0022-5193\(05\)80104-3](https://doi.org/10.1016/S0022-5193(05)80104-3)
- [62] Romanovsky, Y.E. (1985) Food limitation and life-history strategies in cladoceran crustaceans. *Archiv fuer Hydrobiologie*, 21, 363–372.
- [63] Schwenk, K., Posada, D. & Hebert, P.D.N. (2000) Molecular systematics of European *Hyalodaphnia*: the role of contemporary hybridization in ancient species. *Proceedings of the Royal Society B: Biological Sciences*, 267, 1833–1842. <https://doi.org/10.1098/rspb.2000.1218>
- [64] Sharma, P. & Kotov, A.A. (2015) Establishing of *Chydorus sphaericus* (O.F. Muller) s. str. (Crustacea: Cladocera) in Australia: consequences of mass fish stock from Northern Europe? *Journal of Limnology*, 74, 225–233.
- [65] Shiel, R.J. & Dickson, J.A. (1995) Cladocera recoded from Australia. *Transactions of the Royal Society of South Australia*, 119, 29–40.
- [66] Shiel, R.J. & Tan, L.W. (2013) Zooplankton response monitoring: Lower Lakes, Coorong and Murray Mouth October 2011 - April 2012. Final report to Dept of Env't, Water & Natural Resources, Adelaide, 49 pp. <https://doi.org/10.13140/2.1.4129.3440>
- [67] Sievers, F., Wilm, A., Dineen, D., Gibson, T.J., Karplus, K., Li, W., Lopez, R., McWilliam, H., Remmert, M., Söding, J., Thompson, J.D. & Higgins, D.G. (2011) Fast, scalable generation of high-quality protein multiple sequence alignments using Clustal Omega. *Molecular Systems Biology*, 7, 539. <https://doi.org/10.1038/msb.2011.75>
- [68] Smirnov, N.N. (1995) Check-list of the Australian Cladocera (Crustacea). *Arthropoda Selecta*, 4 (1), 3–6.
- [69] Smirnov, N.N. & Timms, B.V. (1983) A revision of the Australian Cladocera (Crustacea). *Records of the Australian Museum*, 1 (Supplement), 1–132. <https://doi.org/10.3853/j.0812-7387.1.1983.103>
- [70] Spaak, P., Fox, J. & Hairston, N.G. (2012) Modes and mechanisms of a *Daphnia* invasion. *Proceedings of the Royal Society of London B: Biological Sciences*, 279, 2936–2944. <https://doi.org/10.1098/rspb.2012.0280>
- [71] Straile, D. & Geller, W. (1998) Crustacean zooplankton in Lake Constance from 1920 to 1995: Response to eutrophication and re-oligotrophication. *Advances in Limnology*, 53, 255–274.
- [72] Swofford, D. (2003) PAUP*. Phylogenetic Analysis Using Parsimony (* and other methods). Version 4. Sinauer Associates, Sunderland (MA). [software]
- [73] Taylor, D.J., Finston, T.L. & Hebert, P.D.N. (1998) Biogeography of a widespread freshwater crustacean: Pseudocongruence and cryptic endemism in the North American *Daphnia laevis* complex. *Evolution*, 52, 1648–1670. <https://doi.org/10.1111/j.1558-5646.1998.tb02245.x>
- [74] Taylor, D.J. & Hebert, P.D.N. (1993) Cryptic intercontinental hybridization in *Daphnia* (Crustacea): the ghost of introductions past. *Proceedings of the Royal Society of London, Series B - Biological Sciences*, 254, 163–168. <https://doi.org/10.1098/rspb.1993.0141>

- [75] Taylor, D.J., Hebert, P.D.N. & Colbourne, J.K. (1996) Phylogenetics and evolution of the *Daphnia longispina* group (Crustacea) based on 12S rDNA sequence and allozyme variation. *Molecular Phylogenetics and Evolution*, 5, 495-510. <https://doi.org/10.1006/mpev.1996.0045>
- [76] Tokishita, S., Shibuya, H., Kobayashi, T., Sakamoto, M., Ha, J.Y., Yokobori, S., Yamagata, H. & Hanazato, T. (2017) Diversification of mitochondrial genome of *Daphnia galeata* (Cladocera, Crustacea): Comparison with phylogenetic consideration of the complete sequences of clones isolated from five lakes in Japan. *Gene*, 611, 38-46. <https://doi.org/10.1016/j.gene.2017.02.019>
- [77] Vaidya, G., Lohman, D.J. & Meier, R. (2011) SequenceMatrix: concatenation software for the fast assembly of multi-gene datasets with character set and codon information. *Cladistics*, 27, 171-180. <https://doi.org/10.1111/j.1096-0031.2010.00329.x>
- [78] Wikipedia (2016) Invasive species in Australia, 2016. Available from: https://en.wikipedia.org/wiki/Invasive_species_in_Australia (accessed 24 September 2017)
- [79] Wooller, I. (2013) Variation in expression of duplicated insulin-like receptor genes in *Daphnia pulex*. Office of Graduate Studies and Research, Central Washington University, Ellensburg, Washington, 84 pp.
- [80] Xu, M., Zhang, H.J., Deng, D.G., Wang, W.-P., Zhang, X.-L., Zha, L.-S. (2014) Phylogenetic relationship and taxonomic status of four *Daphnia* species based on 16S rDNA and COI sequence. *Acta Hydrobiologica Sinica*, 38, 1040-1046.
- [81] Ye, Q., Giatas, G., Aldridge, K., Busch, B., Gibbs, M., Hipsey, M., Lorenz, Z., Mass, R., Oliver, R., Shiel, R., Woodhead, J. & Zampatti, B. (2017) Long-term intervention monitoring of the ecological responses to Commonwealth Environmental Water delivered to the Lower Murray River Selected Area in 2015/16. A report prepared for the Commonwealth Environmental Water Office. South Australian Research and Development Institute, Aquatic Sciences, 184 pp.
- [82] Zuykova, E.I., Bochkarev, N.A. & Katokhin, A.V. (2013) Identification of the *Daphnia* species (Crustacea: Cladocera) in the lakes of the Ob and Yenisei River basins: morphological and molecular phylogenetic approaches. *Hydrobiologia*, 715, 135-150. <https://doi.org/10.1007/s10750-012-1423-3>
- [83] Zuykova, E.I., Bochkarev, N.A., Semenova, A.S. & Katokhin, A.V. (2010) Morphological differentiation, mitochondrial and Nuclear DNA variability between geographically distant populations of *Daphnia galeata* and *Daphnia cucullata* (Anomopoda, Daphniidae). *Journal of Siberian Federal University, Biology*, 4, 434-453.
- [84] Zuykova, E.I., Bochkarev, N.A. & Sheveleva, N.G. (2016) Genetic polymorphism, haplotype distribution, and phylogeny of *Daphnia* (Cladocera: Anomopoda) species from the water bodies of Russia as inferred from the 16S mtDNA gene sequencing. *Russian Journal of Genetics*, 52, 585-596. <https://doi.org/10.1134/S102279541604013X>